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**Term:**

L7 same phosphoramidate

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<u>L8</u>	L7 same phosphoramidate	1	<u>L8</u>
<u>L7</u>	Group same I same intron	471	<u>L7</u>
<u>L6</u>	I5 same exon	1	<u>L6</u>
<u>L5</u>	L2 same Group same I	24	<u>L5</u>
<u>L4</u>	L2 same GroupI	0	<u>L4</u>
<u>L3</u>	L2 same oligonucleotide	1	<u>L3</u>
<u>L2</u>	L1 same intron	36	<u>L2</u>
<u>L1</u>	inhibit\$ same (self near0 splic\$)	55	<u>L1</u>

END OF SEARCH HISTORY

(FILE 'HOME' ENTERED AT 12:14:46 ON 23 OCT 2003)

FILE 'MEDLINE, BIOSIS, CAPLUS, EMBASE' ENTERED AT 12:15:15 ON 23 OCT 2003

L1 2273 S GROUP(W)I(W)INTRON  
L2 229 S L1(P)INHIBIT?  
L3 139 S L2 (P)(SELF(W)SPLIC?)  
L4 10 S L3(P)PHOSPHORAMIDATE  
L5 4 DUPLICATE REMOVE L4 (6 DUPLICATES REMOVED)

=> s l3(p)oligonucleotide

L6 10 L3(P) OLIGONUCLEOTIDE

=> duplicate remove l6

DUPLICATE PREFERENCE IS 'MEDLINE, BIOSIS, CAPLUS, EMBASE'

KEEP DUPLICATES FROM MORE THAN ONE FILE? Y/(N):n

PROCESSING COMPLETED FOR L6

L7 4 DUPLICATE REMOVE L6 (6 DUPLICATES REMOVED)

=> d bib ab l7 1-4

L7 ANSWER 1 OF 4 CAPLUS COPYRIGHT 2003 ACS on STN  
AN 2003:432135 CAPLUS  
TI Folding and targeting RNA  
AU Turner, Douglas; Schroeder, S. J.; Mathews, D. H.; Disney, M. D.; Childs, J. I.  
CS University of Rochester, Rochester, NY, USA  
SO Abstracts, 31st Northeast Regional Meeting of the American Chemical Society, Saratoga Springs, NY, United States, June 15-18 (2003), 50  
Publisher: American Chemical Society, Washington, D. C.  
CODEN: 69EBFV  
DT Conference; Meeting Abstract  
LA English  
AB The databases of genome sequences provide a foundation for developing databases of RNA secondary structures. Free energy minimization with nearest neighbor parameters provides useful predictions of possible secondary structures for a given RNA. These predictions depend on free energy parameters for RNA loops. Recent expts. with loops suggest revisions to models used to approx. these free energies. Predictions of secondary structure are improved by simultaneously minimizing free energy and aligning base pairs for two sequences with identical function. Evidently, this combination partially compensates for incomplete knowledge of RNA thermodyn. A computer program, called Dynalign, automates this combination of free energy minimization and sequence comparison. Predictions of secondary structure can facilitate design of therapeutics that target RNA. For example, the predicted proclivity of RNA to misfold can be exploited by **oligonucleotide** directed misfolding of RNA (ODMiR) strategies to trap an RNA in a nonfunctional conformation. An application to **inhibition** of **self-splicing** by the **group I intron** from *Candida albicans* has been demonstrated.

L7 ANSWER 2 OF 4 MEDLINE on STN DUPLICATE 1  
AN 2001277619 MEDLINE  
DN 21264135 PubMed ID: 11371215  
TI Binding enhancement by tertiary interactions and suicide inhibition of a *Candida albicans* group I intron by phosphoramidate and 2'-O-methyl hexanucleotides.  
AU Disney M D; Matray T; Gryaznov S M; Turner D H  
CS Departments of Chemistry and Pediatrics, University of Rochester, Rochester, New York 14627-0216, USA.  
NC AI45398 (NIAID)  
SO BIOCHEMISTRY, (2001 May 29) 40 (21) 6520-6.  
Journal code: 0370623. ISSN: 0006-2960.

CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals  
EM 200108  
ED Entered STN: 20010820  
Last Updated on STN: 20010820  
Entered Medline: 20010816

AB Candida albicans is one of many infectious pathogens that are evolving resistance to current treatments. RNAs provide a large class of targets for new therapeutics for fighting these organisms. One strategy for targeting RNAs uses short oligonucleotides that exhibit binding enhancement by tertiary interactions in addition to Watson-Crick pairing. A potential RNA target in C. albicans is the **self-splicing group I intron** in the LSU rRNA precursor. The recognition elements that align the 5' exon splice site for a ribozyme derived from this precursor are complex [Disney, M. D., Haidaris, C. G., and Turner, D. H. (2001) Biochemistry 40, 6507-6519]. These recognition elements have been used to guide design of hexanucleotide mimics of the 5' exon that have backbones modified for nuclease stability. These hexanucleotides bind as much as 100000-fold more tightly to a ribozyme derived from the intron than to a hexanucleotide mimic of the intron's internal guide sequence, r(GGAGGC). Several of these oligonucleotides **inhibit precursor self-splicing** via a suicide **inhibition** mechanism. The most promising suicide **inhibitor** is the ribophosphoramidate rn(GCCUC)rU, which forms more trans-spliced than cis-spliced product at **oligonucleotide** concentrations of >100 nM at 1 mM Mg(2+). The results indicate that short oligonucleotides modified for nuclease stability can target catalytic RNAs when the elements of tertiary interactions are complex.

L7 ANSWER 3 OF 4 CAPLUS COPYRIGHT 2003 ACS on STN  
AN 2000:666925 CAPLUS  
DN 133:248036  
TI IGS-binding, phosphoramidate- or thiophosphoramidate-linked oligonucleotides for inhibition of Group I intron self-splicing  
IN Testa, Stephen M.; Disney, Matthew D.; Gryaznov, Sergei M.; Turner, Douglas H.  
PA Geron Corp., USA; University of Rochester  
SO PCT Int. Appl., 37 pp.  
CODEN: PIXXD2

DT Patent  
LA English

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2000055374	A1	20000921	WO 2000-US7045	20000315
	W:	AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW,			
	AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW:	GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG			

PRAI US 1999-124451P P 19990315

AB A method of **inhibiting the self-splicing** of a **Group I intron** is disclosed. The method uses an **oligonucleotide** having a sequence essentially identical to a guide sequence found in the 5' flanking exon and terminates with a 3' ribonucleoside. Usually the **oligonucleotide** has N3'.fwdarw.P5'

phosphoramidate or thiophosphoramidate linkages rather than phosphodiester linkages. A method of **inhibiting** the growth of organisms having **Group I intron**, particularly certain pathogenic fungi including *Pneumocystis carinii*, *Candida albicans* and *Aspergillus nidulans* using the **oligonucleotide** is also provided.

RE.CNT 3        THERE ARE 3 CITED REFERENCES AVAILABLE FOR THIS RECORD  
              ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7    ANSWER 4 OF 4        MEDLINE on STN                        DUPLICATE 2  
AN    2001076965        MEDLINE  
DN    20541381        PubMed ID: 11087376  
TI    Contributions of individual nucleotides to tertiary binding of substrate  
      by a *Pneumocystis carinii* group I intron.  
AU    Disney M D; Gryaznov S M; Turner D H  
CS    Department of Chemistry, University of Rochester, Rochester, New York  
      14627-0216, USA.  
NC    AI45398 (NIAID)  
SO    BIOCHEMISTRY, (2000 Nov 21) 39 (46) 14269-78.  
      Journal code: 0370623. ISSN: 0006-2960.  
CY    United States  
DT    Journal; Article; (JOURNAL ARTICLE)  
LA    English  
FS    Priority Journals  
EM    200101  
ED    Entered STN: 20010322  
      Last Updated on STN: 20010322  
      Entered Medline: 20010111  
AB    *Pneumocystis carinii* is a mammalian pathogen that infects and kills  
      immunocompromised hosts such as cancer and AIDS patients. The LSU rRNA  
      precursor of *P. carinii* contains a conserved **group I**  
      **intron** that is an attractive drug target because humans do not  
      contain group I introns. The **oligonucleotide** r(AUGACU), whose  
      sequence mimics the 3'-end of the 5'-exon, binds to a ribozyme derived  
      from the intron with a  $K(d)$  of 5.2 nM, which is 61000-fold tighter than  
      expected from base-pairing alone [Testa, S. M., Haidaris, G. C.,  
      Gigliotti, F., and Turner, D. H. (1997) *Biochemistry* 36, 9379-9385].  
      Thus, **oligonucleotide** binding is enhanced by tertiary  
      interactions. To localize interactions that give rise to this tertiary  
      stability, binding to the ribozyme has been measured as a function of  
      **oligonucleotide** length and sequence. The results indicate that  
      4.3 kcal/mol of tertiary stability is due to a G.U pair that forms at the  
      intron's splice junction. Eliminating nucleotides at the 5'-end of  
      r(AUGACU) does not affect intron binding more than expected from  
      differences in base-pairing until r((\_\_\_\_)ACU), which binds much more  
      tightly than expected. Adding a C at the 5'- or 3'-end that can  
      potentially form a C-G pair with the target has little effect on binding  
      affinity. Truncated oligonucleotides were tested for their ability to  
      **inhibit** intron **self-splicing** via a suicide  
      **inhibition** mechanism. The tetramer, r((\_\_\_\_)GACU), retains similar  
      binding affinity and reactivity as the hexamer, r(AUGACU). Thus  
      oligonucleotides as short as tetramers might serve as therapeutics that  
      can use a suicide **inhibition** mechanism to **inhibit**  
      **self-splicing**. Results with a phosphoramidate tetramer  
      and thiophosphoramidate hexamer indicate that oligonucleotides with  
      backbones stable to nuclease digestion retain favorable binding and  
      reactivity properties.

**WEST****End of Result Set**

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L8: Entry 1 of 1

File: USPT

Aug 19, 2003

DOCUMENT-IDENTIFIER: US 6608036 B1

TITLE: Oligonucleotide N3'.fwdarw.P5' thiophosphoramidates: their synthesis and administration to treat neoplasms

Other Reference Publication (12):

Testa, S., et al., "In Vitro Suicide Inhibition of Self-Splicing of a Group I Intron from *Pneumocystis Carinii* by An N3'.fwdarw.P5' Phosphoramidate Hexanucleotide", Proc. Natl. Acad. Sci. USA, 96:2734 (Mar., 1999).

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